

Mono- and Bisquinoline-Annulated Porphyrins from Porphyrin β,β' -Dione Oximes

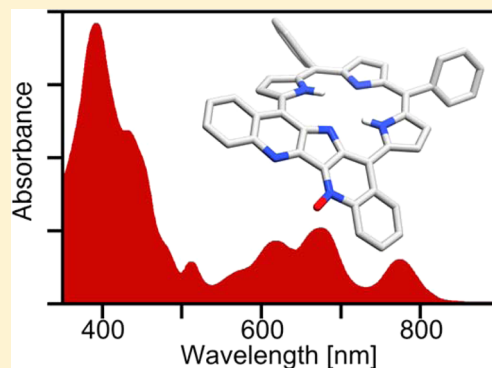
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S Supporting Information

ABSTRACT: An acid-induced reaction of *meso*-tetraphenyl-2-hydroxyimino-3-oxoporphyrin leads, with concomitant loss of water, to a formal electrophilic aromatic substitution of the ortho-position of the phenyl group adjacent to the oxime, forming a quinoline moiety. Owing in part to the presence of a π -extended chromophore, the resulting *meso*-triphenylmonoquinoline-annulated porphyrin ($\lambda_{\text{max}} = 750$ nm) possesses a much altered optical spectrum from that of the starting oxime ($\lambda_{\text{max}} = 667$ nm). An oxidative DDQ-induced ring-closure process is also possible, generating the corresponding *meso*-triphenylmonoquinoline-annulated porphyrin quinoline *N*-oxide, possessing a slightly shifted and sharpened UV-vis spectrum ($\lambda_{\text{max}} = 737$ nm). The connectivity of the chromophores was conclusively shown by NMR spectroscopy. Both ketone functionalities in *meso*-tetraphenyl-2,3-dioxoporphyrin can be converted, via the oxime and using the acid- or oxidant-induced reaction pathways, either in one step or in a stepwise fashion, to bisquinoline-annulated porphyrin ($\lambda_{\text{max}} = 775$ nm) and its *N*-oxide ($\lambda_{\text{max}} = 779$ nm), respectively. This process is complementary to a previously established pathway toward bisquinoline-annulated porphyrins. Their zinc(II), nickel(II), and palladium(II) complexes are also described. Several examples of the quinoline-annulated porphyrins were crystallographically characterized, proving their connectivity and showing their conformations that are extremely distorted from planarity. The work presents a full account on the synthesis, structure, and spectroscopic properties of these classes of NIR-absorbing dyes.



■ INTRODUCTION

Even though the high extinction coefficients, fluorescence, and singlet oxygen photosensitization quantum yields of regular porphyrins and chlorins are potentially attractive for their use in biomedical applications, e.g., as photochemotherapeutics,¹ photoantimicrobials,² photoantifungals,³ or fluorescent markers,^{1c,4} most regular porphyrins and chlorins do not absorb light within the ~ 700 – 900 nm wavelength regime referred to as the “optical window” of tissue.⁵ For instance, the maximum wavelength of absorbance (λ_{max}) for regular porphyrins, such as *meso*-tetraphenylporphyrin **1H₂**, rarely exceeds 650 nm. In comparison, the wavelength of maximum penetration of breast tissue is ~ 725 nm; whole blood has an absorption minimum at ~ 710 nm.⁶ Regular porphyrins are thus unsuitable for biomedical *in vivo* applications. Nonemitting chromophores may also be attractive as photoacoustic imaging agents but,⁸ again, only if they also absorb light within the optical window of tissue.⁹ Light-harvesting applications also benefit from NIR-absorbing dyes, as a large portion of the irradiance energy of sunlight falls into this range. Panchromatic absorbers are also most desirable for light harvesting, as these chromophores collect the maximum amount of energy.⁷

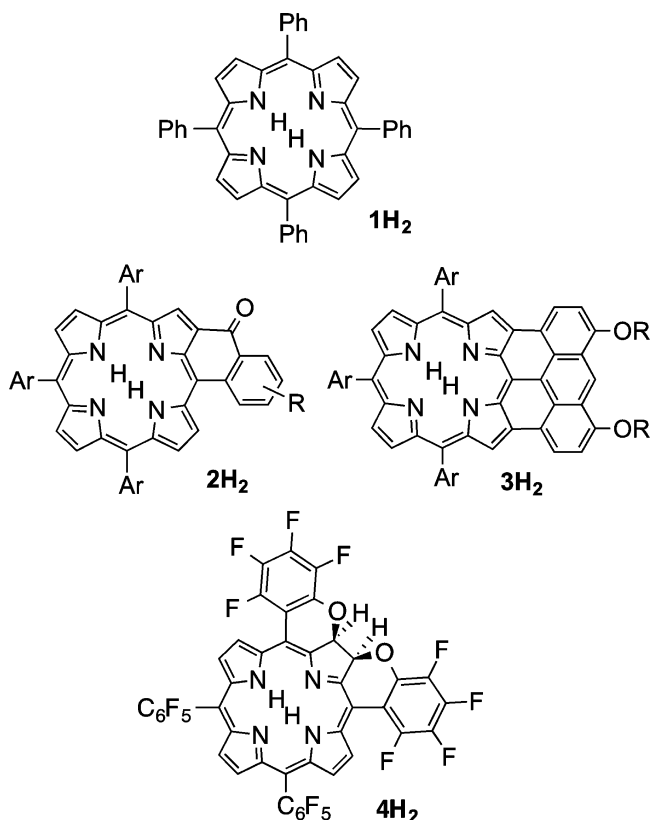
A number of strategies have been employed to achieve this goal. For instance, the expansion of the conjugated π -system by increasing the number of conjugated pyrroles was greatly

successful, as the many examples of NIR-absorbing expanded porphyrins demonstrate.¹⁰ One other strategy is the establishment of π -systems that are annulated to the porphyrinic chromophore.^{7d,11} Among the latter class of porphyrins may be *meso*-arylporphyrin derivatives that bear a covalent linkage between one or more β -positions and the flanking *meso*-phenyl/aryl groups.¹² In the absence of the linkage, a $H_{\beta\text{-pyrrole}} - H_{\alpha\text{-phenyl}}$ steric interaction prevents the *meso*-aryl groups from adopting low-energy coplanar conformations. The linkage removes this interaction but needs to be short enough to force the phenyl group(s) into (idealized) coplanarity with the porphyrinic chromophore, thereby extending the π -conjugation pathway. This linkage may be a ketone functionality, itself in conjugation with the chromophore (**2H₂**).¹³ The resulting bathochromic shift of λ_{max} of **2H₂** compared to **1H₂** is a respectable ~ 76 nm. On the other hand, the fusion of one through four anthracenes to a porphyrin, as in **3H₂**, results in an enormous perturbation of their UV-vis spectra, with λ_{max} values red-shifted ~ 300 nm upon fusion of a single anthracene; the corresponding nickel(II) complex with four anthracenes shifted the λ_{max} by more than 900 nm, to 1417 nm.¹⁴ Few examples exist in which a chlorin chromophore incorporates an

Received: November 2, 2014

Published: December 3, 2014

annulated ring, as in bischromene-annulated chlorin **4H₂**, leading to a 24 nm-shifted λ_{max} compared to the parent chlorin.¹⁵



In 2011, Jeandon and Ruppert, building on the extensive work by Ruppert, Callot, and co-workers in the construction of peripherally conjugated chelates,¹⁶ published the formation of bisquinoline-annulated porphyrin **7H₂** from nickel porphyrin **1Ni** via a multistep sequence (Scheme 1).¹⁷ The key β -to-*ortho*-linkage formation step involved a Cadogan reaction of

nitroporphyrin **5Ni** to form the amine-linked porphyrin **6Ni**. Subsequent nitrogen protection, regioselective nitration, thermally induced oxidation and ring-closure, and acid-induced demetalation produced the bisquinoline-annulated system **7H₂** in good yield.

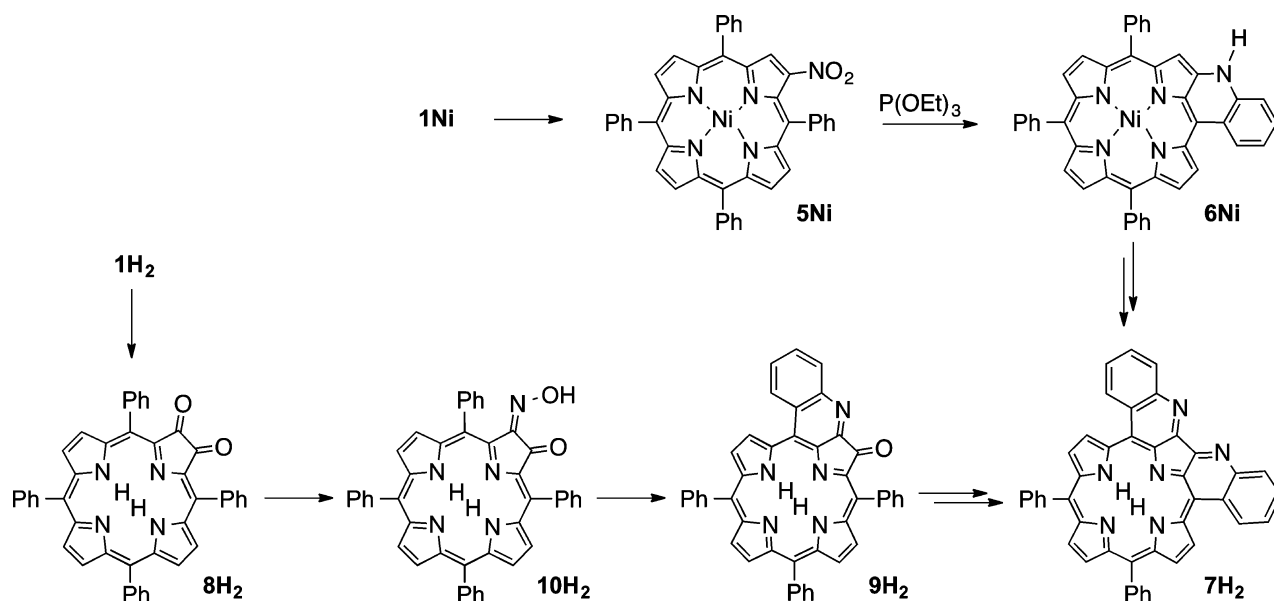
Concurrently with Jeandon and Ruppert, we independently described in a preliminary report the conversion of free base porphyrin **1H₂** to free base bisquinoline-annulated porphyrin **7H₂**.¹⁸ The key steps were two consecutive acid-mediated electrophilic aromatic substitution reactions of oximes, such as **10H₂**, formed from the corresponding ketone **8H₂**. We prepared this well-known dione **8H₂** along a dihydroxylation–diol oxidation sequence from porphyrin **1H₂**,¹⁹ but this product can be made along a number of alternative routes.²⁰

We follow up here on our preliminary report and present the full account on the formation and reactivity of the mono- and dioximes of **8H₂** with respect to their conversion to a number of free base and metalated mono- and bisquinoline-annulated porphyrins, and their *N*-oxide derivatives. These chromophores are characterized by remarkably bathochromically shifted optical spectra when compared to regular porphyrins. We will also report the X-ray crystal structures of a number of derivatives, thus highlighting their nonplanar conformations we interpret as one reason for their red-shifted and broadened spectra. We recently demonstrated the efficacy of the quinoline-annulated porphyrins to act as photoacoustic imaging contrast agents in tissue phantom studies.⁹

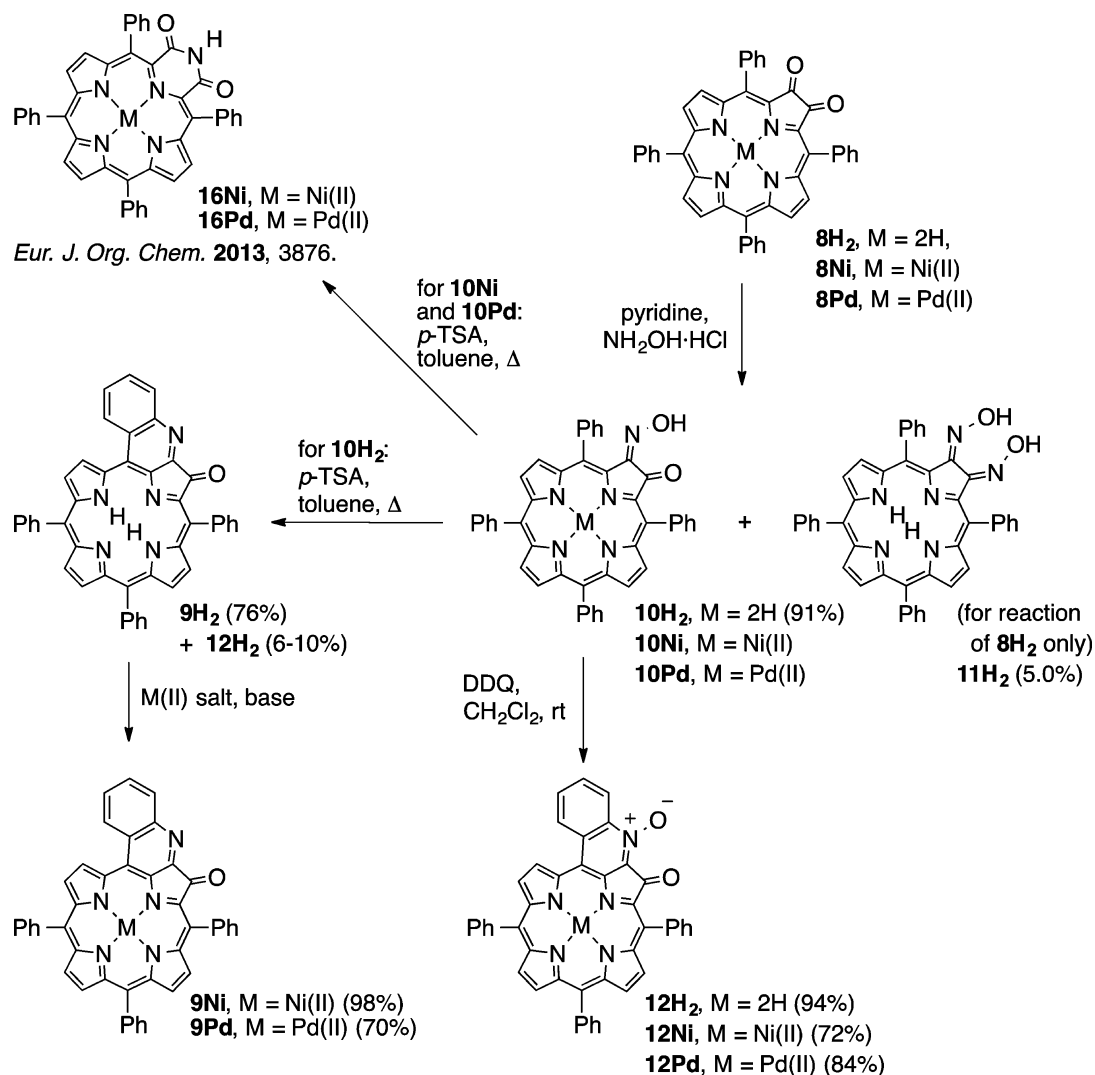
RESULTS AND DISCUSSION

Synthesis of meso-Tetraphenylmonooxime 10H₂ and -bisoxime 11H₂. Analogous to the previously described conversion of the nickel(II) and palladium(II) complexes of porphyrin dione **8Ni** and **8Pd**,²¹ free base 2,3-dioxoporphyrin **8H₂** can also be converted to the corresponding monooxime **10H₂** by reaction with ~100-fold stoichiometric excess of hydroxylamine hydrochloride (NH₂OH·HCl) in pyridine over 24 h at ambient temperature (Scheme 2). Its analytical and spectroscopic properties were similar to those of its

Scheme 1. Two Independent Paths toward Bisquinoline-Annulated Porphyrin 7H₂ Developed by Ruppert (1Ni → 5Ni → 7H₂) and Brückner (1H₂ → 10H₂ → 7H₂)



Scheme 2. Synthetic Pathways toward Monoquinoline-Annulated Porphyrins



diamagnetic metal complexes: the ^{13}C NMR of 10H_2 indicated the presence of one carbonyl (188.2 ppm) and one imine carbon (151.8 ppm), functionalities that could also be confirmed by FT-IR spectroscopy ($\nu_{\text{C=O}}$ and $\nu_{\text{C=N}}$ at 1652.1 and 1548.6 cm^{-1} , respectively). A diagnostic ^1H NMR spectral feature for the monooximes is the signal at ~ 15.7 ppm (exchangeable with D_2O) that is assigned to the oxime hydrogen that is H-bonded to the neighboring carbonyl group (Figure 1). The structural characterization of 10Ni by single crystal X-ray diffractometry is described below (Figure 4).

Next to the major product monooxime 10H_2 , a minor amount of the bisoxime 11H_2 could also be isolated, possessing all expected spectroscopic properties (see Supporting Information (SI) for details). The occurrence of the bisoxime was not observed in the reaction of the dione metal complexes 8Ni and 8Pd .²¹

The UV–vis spectra of free base dione 8H_2 , monooxime 10H_2 , and bisoxime 11H_2 are similar to each other in that all possess, compared to the regular porphyrin spectrum of *meso*-tetraphenylporphyrin 1H_2 (not shown), slightly broadened Soret bands and very broadened Q-bands (Figure 2). However, with increasing ketone-to-oxime transformations, the Q-band region becomes more defined, with increasingly sharper and

distinctive peaks. The strong electronic effects of the β,β' -diketone functionality were discussed previously.^{19,22} Evidently, this influence extends also to the oximes, albeit to a slightly lesser degree.

Formation of Free Base and Metalated Monoquinoline-Annulated Oxoporphyrins 9. Treatment of the olive-colored free base monooxime 10H_2 with a strong organic acid ($p\text{-TSA}$) under forcing conditions (toluene, reflux) generated a major light brown product, 9H_2 , with a mass indicating the loss of H_2O (HR-MS ESI+, 100% CH_3CN , suggests a composition of $\text{C}_{44}\text{H}_{28}\text{N}_3\text{O}$ for MH^+) (Scheme 2). As the oxime signal vanished from the ^1H NMR spectrum of the product, the oxygen lost likely stems from the oxime functionality (Figure 1). Further, the ^1H NMR spectrum of product 9H_2 lacks any axial symmetry: For instance, all six β -protons are non-equivalent. A diagnostic set of four peaks assigned to one phenyl group (confirmed by 2D NMRs; see SI) suggests establishment of a link between a phenyl group and a flanking β -position. Its proton signals are much shifted (between 9.0 and 7.6 ppm) because of the induced coplanarity of this phenyl group with the porphyrinic macrocycle. Indaphyrins, for example, have shown similarly diagnostic patterns for such β -to-*o*-phenyl linkages. The ^{13}C NMR and IR (neat) spectra of 9H_2 indicated the preservation of one β -imine and one β -

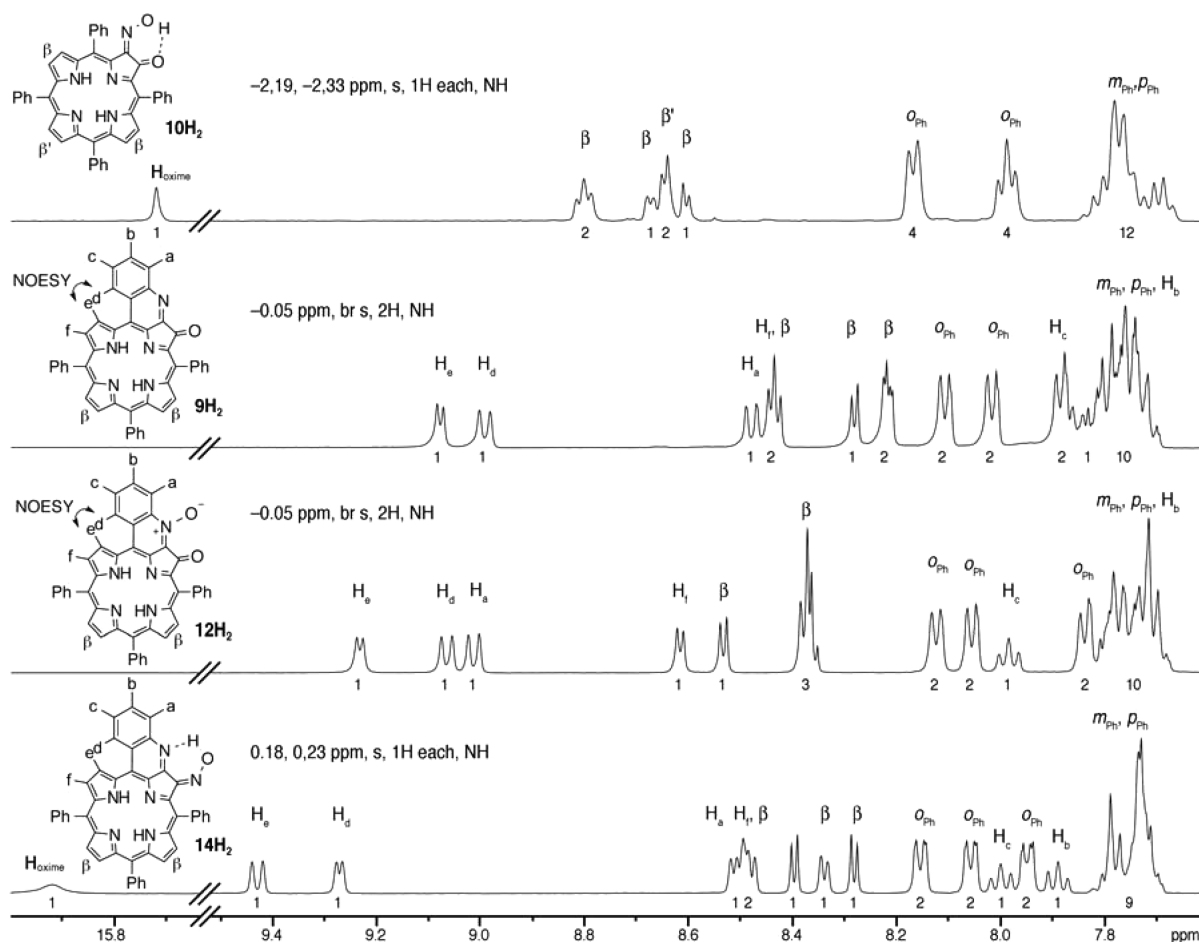


Figure 1. Comparison of the low-field region of the ^1H NMR spectra (400 MHz, CDCl_3 , 25 $^\circ\text{C}$) of the compounds indicated.

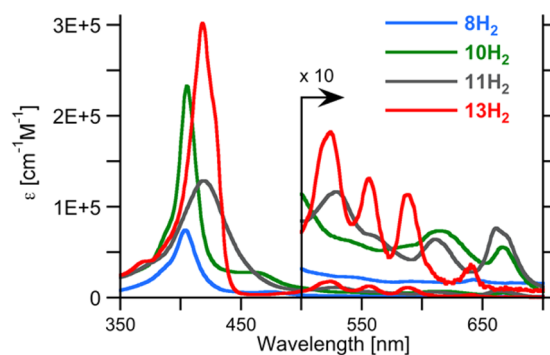


Figure 2. UV-vis spectra (CH_2Cl_2) of the compounds indicated.

ketone carbon (at 151 and 196 ppm, respectively). The summation of the spectroscopic data is suggestive of formation of the quinoline-annulated structure shown in Scheme 2. The UV-vis spectroscopic properties of this compound are, together with the optical spectra of all the other quinoline-annulated porphyrins, described below.

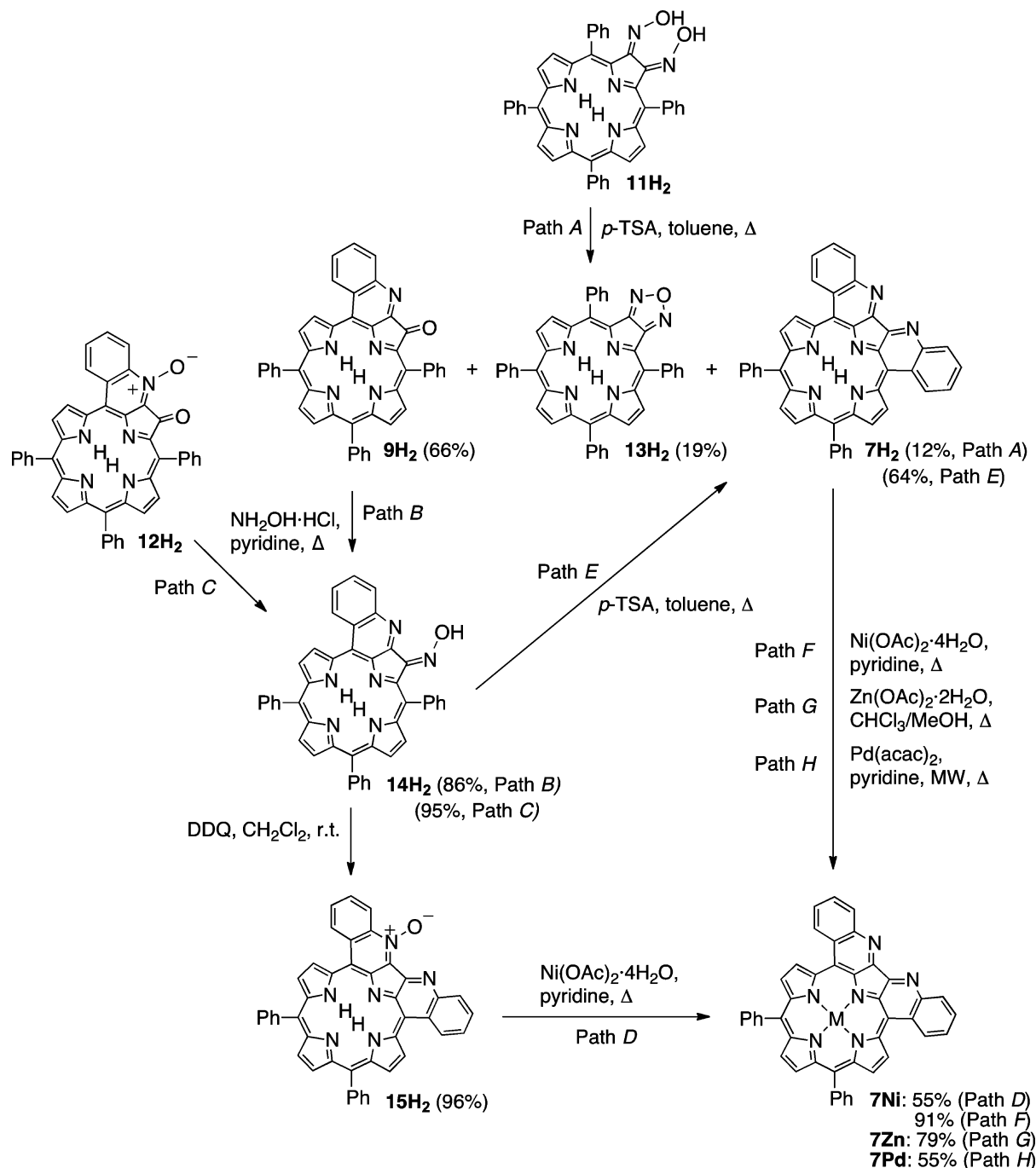
This outcome of the reaction highlights an interesting directing effect of the coordinated metal ion in light of the fact that the metal complexes of monooxime **10Ni** and **10Pd**, when reacted under identical acidic conditions, undergo a Beckmann rearrangement to form the corresponding *meso*-phenylpyrazinoporphyrimetal complexes **16M**.²¹ This reactivity further implies that metal complexes of the quinoline-annulated porphyrin **9M** needed to be prepared by metal insertion into

free base **9H₂**. The corresponding metal insertion reactions using standard methodologies proceeded smoothly with good to excellent yields.²³ The composition (as determined by DART+ HR-MS) and spectroscopic data of **9Ni** and **9Pd** confirmed their expected quinoline-annulated porphyrin structures.

Alongside the major product **9H₂**, we also isolated a minor product **12H₂** in yields ranging from 6% to 10%. Its mass, as determined by HR-MS, indicated the presence of an additional oxygen atom. Performance of the ring-closing reaction under inert conditions largely suppressed its formation. Consequently, the formation of this novel chromophore was traced to air oxidation. Product **12H₂** is also accessible as the major product from free base monooxime **10H₂** under oxidizing conditions.

Synthesis of Monoquinoline-Annulated Oxoporphyrin N-Oxides 12. Guided by the discovery of the oxidized quinoline-annulated product during the acid-catalyzed reaction, we discovered that this product can be prepared as the main product when free base monooxime **10H₂** is treated with DDQ at ambient temperatures (Scheme 2). Interestingly, the metal complexes of the oxime **10M** ($\text{M} = \text{Ni(II)}, \text{Pd(II)}$) are also susceptible to this conversion, generating the metal analogues of the product, **12M** ($\text{M} = \text{Ni(II)}, \text{Pd(II)}$). A careful NaBH_4 reduction of *N*-oxide **12H₂** regenerates **9H₂**, though DDQ oxidation of **9H₂** does not lead to formation of **12H₂**. This suggests that monoquinoline-annulated chromophore **9H₂** is not an intermediate in the formation of **12H₂**. The oxidation of α,β -oximes of unsaturated substrates under a number of

Scheme 3. Synthetic Pathways toward Bisquinoline-Annulated Porphyrins



conditions frequently results in formation of iminoxy radical intermediates that undergo ring-cyclization reactions producing isoxazoles,²⁴ including isoxazoles annulated to a chlorin framework,²⁵ though the formation of quinoline *N*-oxides was also observed.²⁶ Presumably the formation of a six-membered quinoline ring in the cyclization of 10H₂ directs this reaction.

The presence of the oxygen atom in 12M also has a strong influence on the UV–vis spectra of these chromophores when compared to the quinoline-annulated compounds 9M (Figure S; see discussion below). The NMR spectroscopic data for 12M are significantly different from those of corresponding 9M (Figure 1). Nonetheless, the same number of hydrogens and carbons are observed and the compound possesses the same overall symmetry by ¹H NMR spectroscopy (Figure 1). One

hydrogen signal assigned to the CH group adjacent to the quinoline nitrogen is particularly strongly affected by the oxidation, suggestive of the presence of a quinoline *N*-oxide, as opposed to a porphyrin pyrrole *N*-oxide. The latter compounds are known, and their optical spectra are also severely distorted compared to those of the parent porphyrins.²⁷ A crystal structure analysis eventually provided unambiguous proof for the quinoline-annulated porphyrin quinoline *N*-oxide connectivity of product 12M (Figure 4; all crystal structures are discussed below).

Synthesis of Monoquinoline-Annulated Oxoporphyrin Oxime 14H₂. Treatment of the β -keto functionality of free base monoquinoline 9H₂ with hydroxylamine hydrochloride under similar conditions as described above for the formation

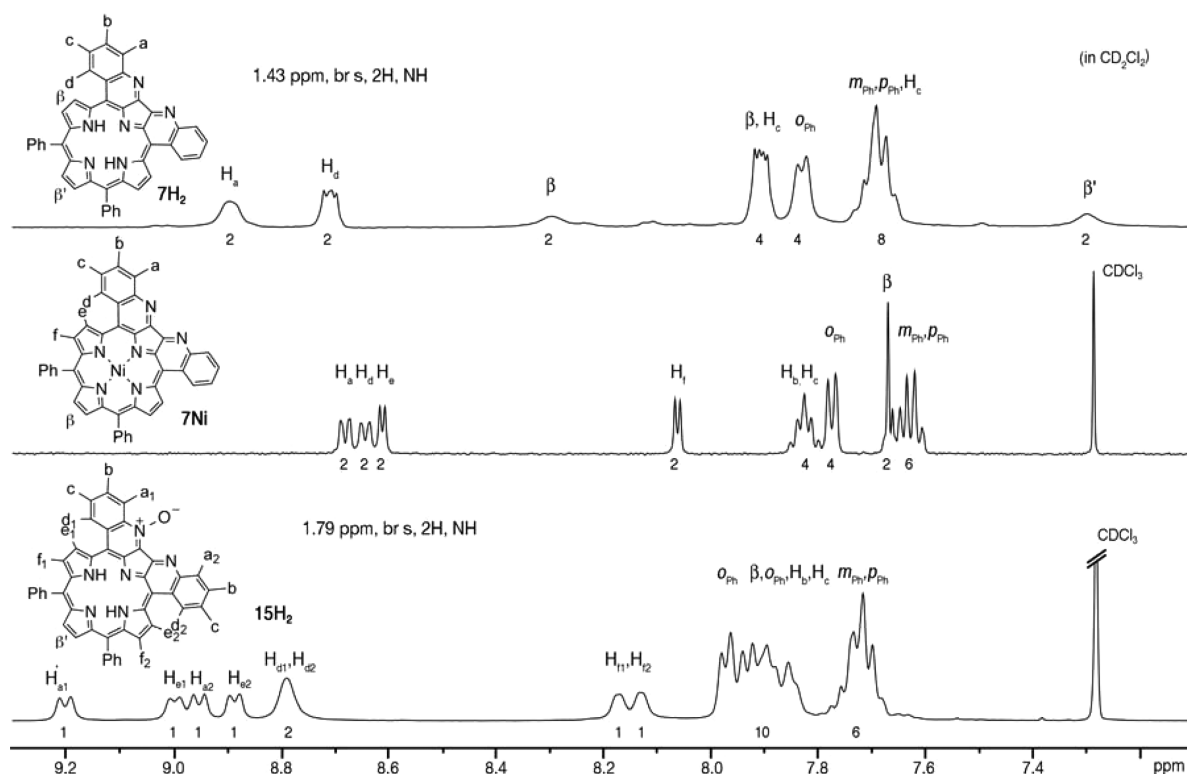


Figure 3. Comparison of the low-field region of the ^1H NMR spectra (400 MHz, in solvents indicated, 25 °C) of the bisquinoline-annulated porphyrins shown.

of the oximes $10\text{H}_2/10\text{M}$ resulted in formation of oxime 14H_2 (Scheme 3). Similarly, reaction of β -keto N -oxide 12H_2 with hydroxylamine converted it to quinoline oxime 14H_2 . Note that the N -oxide moiety is lost in the process; such (reductive) losses are not unusual for heterocycle N -oxides.²⁸ Diagnostic peaks for the resulting oxime 14H_2 in the ^1H NMR spectrum are, analogous to oximes $10\text{H}_2/10\text{M}$, the signal for the strongly H-bonded oxime hydrogen (at ~ 15.9 ppm) (Figure 1). All other spectroscopic and analytical data are as expected. The β -ketone-to-oxime conversion ($9\text{H}_2/12\text{H}_2 \rightarrow 14\text{H}_2$ conversion) perturbs the UV–visible spectrum of the products much more than the corresponding dione to oxime conversions ($8\text{H}_2 \rightarrow 10\text{H}_2/11\text{H}_2$ conversions, Figure 2; see also below).

Synthesis of Bisquinoline-Annulated Porphyrins $7\text{H}_2/7\text{M}$ and Bisquinoline-Annulated Porphyrin N -Oxide 15H_2 . The oxime functionality shows analogous reactivity in both 10H_2 and the quinoline-annulated porphyrin 14H_2 (Scheme 3). Reaction of oxime 14H_2 with p -TSA under reflux conditions resulted in formation of a product in good yield that we were, based on its spectroscopic properties, able to identify as bisquinoline-annulated porphyrin 7H_2 ; it possessed all the spectroscopic properties as reported by Jeandon and Ruppert,¹⁷ including the broadened NMR spectra at rt attributed to extensive stacking and the limited solubility of this compound (Figure 3). Other annulated porphyrins have been observed to form tightly associated stacks in the crystal lattice.^{14c} The synthetic routes presented here are thus alternative and fully independent routes toward a π -extended porphyrin chromophore with much altered optical properties (discussed below). The two pathways toward 7H_2 are complementary to each other. Both possess distinct advantages and disadvantages. The Jeandon and Ruppert synthesis is somewhat shorter (7 steps) versus our pathway (~ 9 steps, depending in the chosen path

toward the dione 8H_2), and the overall yield of both syntheses are also similar ($\sim 20\%$). One advantage of our route is its flexibility with respect to accessing other quinoline-annulated derivatives (such as the monoquinoline-annulated porphyrins and N -oxides).

The metal ions nickel(II), zinc(II), and palladium(II) could be inserted into free base 7H_2 , forming the corresponding metal complexes in good to satisfying yields. Metal insertion led to the formation of compounds that possess much sharpened ^1H NMR spectra (see Figure 3 for the spectrum of 7Ni), presumably because of a lesser degree of stacking.

Reaction of oxime 14H_2 with DDQ led to a compound of the molecular mass of bisquinoline-annulated porphyrin 7H_2 plus an oxygen atom (as per ESI+ HR-MS). Its ^1H NMR spectrum also shows the loss of axial symmetry (Figure 3). This suggests the formation of bisquinoline N -oxide 15H_2 , an interpretation that could also be confirmed by single crystal X-ray diffractometry (discussed below). Nickel(II) insertion into the N -oxide led to loss of the N -oxide functionality and formation of 7Ni .

A Shorter Route to Bisquinoline-Annulated Porphyrin 7H_2 . The successful stepwise formation of bisquinoline-annulated porphyrin 7H_2 via the route dione $8\text{H}_2 \rightarrow$ oxime $10\text{H}_2 \rightarrow$ quinoline-annulated oxoporphyrin $9\text{H}_2 \rightarrow$ quinoline-annulated oxoporphyrin oxime $14\text{H}_2 \rightarrow 7\text{H}_2$ prompts the question whether the direct conversion of bisoxime $11\text{H}_2 \rightarrow 7\text{H}_2$ can be accomplished (even though the formation of bisoxime 11H_2 is inefficient and could not be optimized in our hands). The answer is yes but at a loss of overall efficiency. Reaction of 11H_2 under the conditions leading to quinoline annulation generated the desired product 7H_2 , but in only 12% yield. The major product is quinoline-annulated oxoporphyrin 9H_2 . Evidently, hydrolysis of one of the oximes is faster than

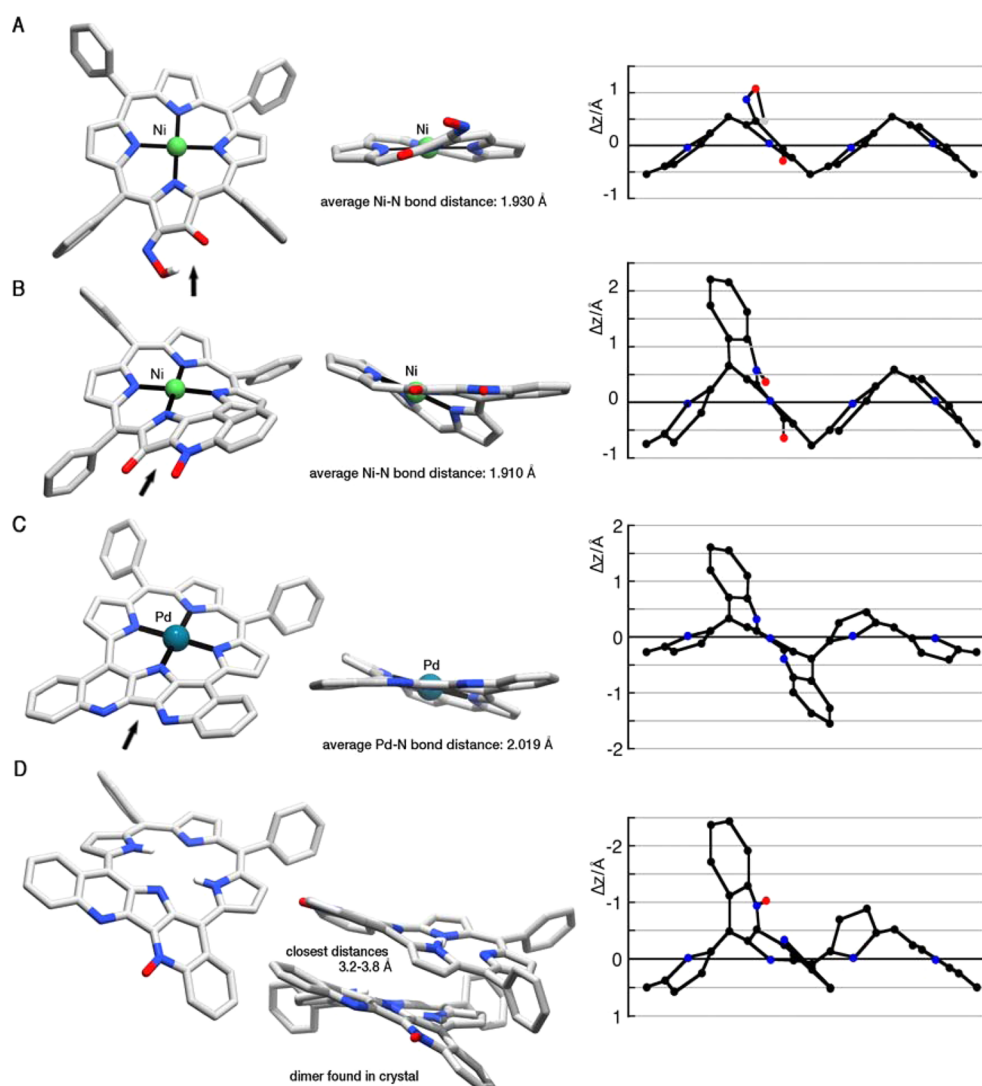


Figure 4. Stick representation of the X-ray single crystal structures: (A) **10Ni**; (B) **12Ni**; (C) **7Pd**; (D) **15H₂**. Top or oblique views (left column), side views (middle column), and deviation of the skeletal heavy atoms from the N₄ mean plane defined by the four pyrrolic nitrogens, as seen in the crystal structures (right column). All disorder and hydrogens bonded to carbon removed for clarity; phenyl groups removed for clarity in A, B, and C. Arrows indicate perspective for side views.

ring-closure. In fact, we observed fast hydrolysis of **11H₂** to form **10H₂**, even under the conditions of silica gel chromatography. The intramolecular condensation of the two oxime moieties with each other is also a more efficient process, generating oxadiazole (furazan)-annulated porphyrin **13H₂**. This novel product, related to Smith's pyrroloporphyrins,²⁹ shows very porphyrin-like optical properties (Figure 2) and was not further investigated (for details of **13H₂**, see also SI). In general, the condensation of α,β -bisoximes resulting in the formation of oxadiazoles is a well-known reaction.³⁰ Thus, even though the stepwise route toward product **11H₂** is significantly longer, its overall yield (~35% from diketone **11H₂**) is much better than the short route via bisoxime **11H₂** (<0.5% from diketone **11H₂**).

Reduction of Monoquinoline-Annulated Porphyrin 9H₂. Treatment of **9H₂** with NaBH₄ in CH₂Cl₂/10% MeOH at ambient temperatures yielded two polar products with a composition (as per ESI⁺-MS) corresponding to [M + 5H]⁺ and another, even more polar product, corresponding to [M + 3H]⁺, suggestive of a reduction of either only the imine double bond (forming one isomer) or both the imine and the ketone

double bonds (forming two diastereomers), respectively. The single double bond reduction product can also be made as the sole product using NaBH₃CN at ice temperatures. Remarkably, all isolated products possessed much more blue-shifted, chlorin-like optical spectra with broadened Soret bands, highlighting the importance of the presence of the ketone and imine functionality for the electronic structure of the chromophore. Much to our disappointment, however, all three products proved to be intractable, as they all rapidly decomposed upon isolation at preparatively useful scales.

Structural Characterization of the Quinoline-Annulated Porphyrins. The considerably ruffled conformation of the monooxime **10Ni** is comparable to that observed in nickel(II) porphyrins and nickel dione **8Ni**¹⁹ and like in these cases can be attributed to the small ionic radius of the low-spin nickel(II) (Figure 4).³¹ The H-bond of the oxime moiety to the adjacent ketone functionality, seen in solution state (cf. to Figure 1), is also expressed in the solid state.

The crystal structure of the nickel(II) complex of quinoline-annulated porphyrin *N*-oxide **12Ni** provided the ultimate proof of the quinoline-annulated porphyrin connectivity, including

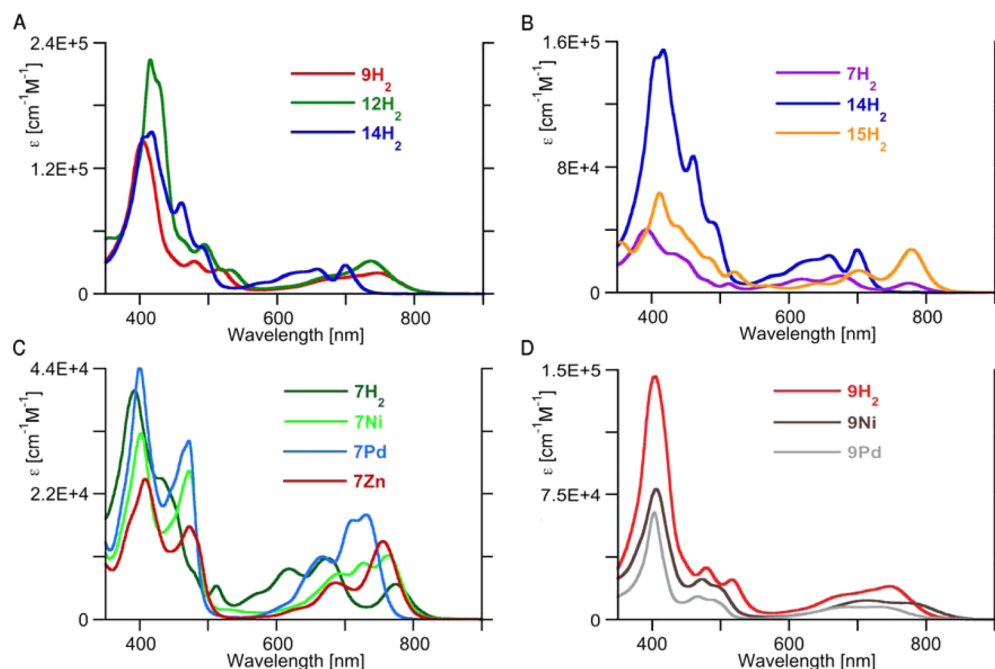


Figure 5. UV-vis spectra of the compounds indicated in CH_2Cl_2 , except compound **7Zn** was, for solubility reasons, recorded in DMSO.

the presence of a quinoline *N*-oxide, as opposed to a central porphyrin pyrrole-*N*-oxide functionality. The ring-closure reaction of the oxime with the neighboring phenyl group generated a pyrrolo[3,2-*b*]quinolin-4-one moiety that is, presumably because of a steric interaction of this moiety with the β -hydrogen next to it, slightly bent. The prime conformation of the porphyrinic macrocycle is still ruffled, but it is strongly modulated by the presence of the quinoline-phenyl-to- β -H interaction. The *N*-oxide and ketone oxygens are eclipsed and form a potential metal binding pocket. Structurally related systems have been used for the formation of external metal complexes.^{16c,d,f,32}

The Ni–N bond distances in primarily ruffled porphyrins can be used to gauge the degree of distortion. In **12Ni**, the Ni–N bond distances average 1.910 Å (ranging from 1.901 to 1.925 Å) and thus fall into the regime of a moderately nonplanar porphyrin. The Ni–N bond lengths in ruffled porphyrins shorten with the degree of ruffling. In comparison, the Ni–N bond lengths in lesser ruffled **10Ni** are 1.930 Å, and in only slightly ruffled **1Ni** 1.931 Å,³³ whereas they are 1.892 Å in an extremely ruffled nickel(II) secochlorin bisaldehyde.^{31c}

The porphyrinoid macrocycle in free base bisquinoline-annulated *N*-oxide **15H₂** is particularly nonplanar, while the pentacyclic annulated heterocycle is slightly and asymmetrically bowed. The presence of palladium in the bisquinoline-annulated porphyrin **7Pd** planarizes the system somewhat when compared to the related free base **15H₂**, but the porphyrinic chromophore is still considerably ruffled with significant saddling and waving modes. Despite these deformations, the metal is coordinated in a distorted square planar fashion, with regular average N–Pd bond distances of 2.019 Å (individual bonds range from 1.983 to 2.041 Å).³³

Optical Properties of Quinoline-Annulated Porphyrins. Upon cyclization of monooxime **10H₂**, the spectrum of quinoline-annulated porphyrin **9H₂** is much altered (Figure 5A): Most obvious, the Q-band region is much bathochromically shifted (for **10H₂**, $\lambda_{\text{max}} = 661$ nm; for **9H₂**, $\lambda_{\text{max}} = 750$ nm). Key to this is believed to be the extension of the

conjugated π -system and the nonplanar conformation of **9H₂** (we extend from the conformation of its *N*-oxide **12H₂**). The latter is also supported by the reduced extinction coefficient for the broadened Soret band of **10H₂**.³⁴ Comparing the spectra of quinoline-annulated porphyrin **9H₂** with that of its *N*-oxide **12H₂**, the distinct and sharpening effect of the quinoline *N*-oxidation ($\lambda_{\text{max}} = 737$ nm) and a red-shift of the Soret region can be noticed, highlighting the strong electronic effect the quinoline moiety has on the porphyrinic chromophore. Conversion of the oxo-group in **9H₂** to an oxime in **14H₂** ($\lambda_{\text{max}} = 700$ nm) similarly has a general sharpening effect on the Q-bands as the conversion of diketone **8H₂** to oxime **10H₂** (cf. to Figure 2).

Cyclization of quinoline-annulated porphyrin monooxime **14H₂** leads to the red-shifted spectrum of bisquinoline porphyrin **7H₂** described previously (**7H₂**: $\lambda_{\text{Soret}} = 392$, $\lambda_{\text{max}} = 775$ nm) (Figure 5B).^{17,18} Again, we attribute this shift to the conformation of the chromophore and the presence of the pentacyclic heterocycle that is π -conjugated with the porphyrinoid ring. As was observed for the oxidation of monoquinoline **9H₂**, *N*-oxidation of **7H₂**, forming **15H₂**, also leads to a red-shift of its Soret band ($\lambda_{\text{Soret}} = 412$, $\lambda_{\text{max}} = 779$ nm). The closed-shell metal complexes of bisquinoline-annulated porphyrin **7H₂**, **7Zn**, **7Ni**, and **7Pd** show, as expected, all similarly patterned spectra with a reduced number of Q-bands and a split Soret band (Figure 5C).

In contrast to the UV-vis spectra of metal complexes of bisquinoline **7H₂**, the UV-vis spectra of the nickel and palladium complexes of monoquinoline porphyrin **9H₂**, **9Ni** and **9Pd**, are not much different from those of the free base spectrum: they possess a much broadened Q-band region, with no sign of a split Soret band (Figure 5D). In this respect they are more similar to the metal complexes of dione **8H₂**.

CONCLUSIONS

We demonstrated the annulation of a quinoline moiety onto a *meso*-tetraphenylporphyrin macrocycle by intramolecular ring

closure of the oximes of the well-known porphyrin 2,3-diones. This strategy can be applied twice in a stepwise fashion or, less efficiently, in a double-ring closure reaction of an α,β -bisoxime. Depending on the reagents used, a condensation reaction to form the quinoline-annulated ring-system takes place, or this condensation is also coupled with an oxidation. Thus, mono- and bisquinoline-annulated porphyrins and their quinolone *N*-oxides became accessible. These classes of porphyrin derivatives possess dramatically bathochromically shifted and broadened UV-vis spectra compared to the spectrum of the fundamental starting material porphyrin **1H₂**. This is attributed to the direct conjugation of the quinoline moiety with the aromatic porphyrinic π -system, the strong influence of β -oxo groups, and the nonplanarity of the chromophore. This synthesis of the bisquinoline-annulated porphyrin is an alternative to the synthesis presented by Jeandon and Ruppert.¹⁷ The solid-state conformations of all quinoline-annulated porphyrins display pronounced deviations from planarity because of increased pyrrole- β -to-*o*-aryl interactions with the ring-annulated systems, while the polycyclic quinoline-annulated part of the compounds itself remains idealized planar or slightly curved. The conformation is modulated by the central metal present. The broad and intense absorption of the chromophores in the NIR region suggests their use in biomedical applications. However, their very low fluorescence and singlet oxygen quantum yields, reported earlier,⁹ prevent their use as PDT agents, but we were able to show that this class of porphyrinoids is suitable as contrast agents for photoacoustic imaging applications.⁹ We are currently in the process of preparing a range of quinoline-annulated porphyrins that possess more favorable solubility properties for their use in aqueous biological environments.

EXPERIMENTAL SECTION

Materials and Instruments. Solvents and reagents were used as received. Aluminum-backed, silica gel 60, 250 μ m thickness analytical plates, 20 \times 20 cm, glass-backed, silica gel 60, 500, or 1000 μ m thickness preparative TLC plates, and standard grade, 60 Å, 32–63 μ m flash column silica gel were used. Diones **8H₂**, **8Ni**, and **8Pd** were prepared as described previously.^{19,21}

X-ray Single Crystal Diffractometry. Single crystals of **10Ni**, **12Ni**, **15H₂**, and **7Pd** were coated in mineral oil, mounted on a micromesh mount, and placed on a goniometer head under a stream of nitrogen cooled to 100 K. The data were collected on diffractometers using monochromatic Mo K α (**15H₂**, **12Ni**, and **10Ni**) or Cu K α radiation (**7Pd**) with the omega scan technique. Data were collected, their unit cells determined, and the data integrated and corrected for absorption and other systematic errors using the Apex2 suite of programs. The frames were integrated with the Bruker SAINT software package using a narrow-frame algorithm. Data were corrected for absorption effects using the multiscan method (SADABS). Space groups were assigned and the structures were solved by direct methods using the SHELXTL suite of programs and were refined by full matrix least-squares against F^2 with all reflections using Shelxl-97 or Shelxl2013 until the final anisotropic full-matrix, least-squares refinement of F^2 converged. Details of the data collection and structural parameters for the structure elucidations of **10Ni**, **12Ni**, **15H₂**, and **7Pd**, including complete cif files, descriptions of disorder, and hydrogen atom treatment, are presented in the SI.

meso-Tetraphenyl-2-hydroxyimino-3-oxoporphyrin (10H₂) and meso-Tetraphenyl-2,3-dihydroxyiminoporphyrin (11H₂). *meso*-Tetraphenyl-2,3-dioxochlorin **8H₂** (120 mg, 1.85×10^{-4} mmol) was dissolved in pyridine (25 mL) in a round-bottom flask equipped with a magnetic stir bar and N₂ gas inlet. Hydroxylamine hydrochloride (H₂NOH·HCl, 1.2 g, \sim 100 equiv) was added, and the mixture was stirred for 24 h at rt. When the starting material was

consumed (reaction control by TLC), the reaction mixture was evaporated to dryness by rotary evaporation, taken up in CH₂Cl₂, and filtered through a glass frit (M). The volume of the filtrate was reduced and the mixture separated by column chromatography (silica-CH₂Cl₂/petroleum ether 3:1), allowing the isolation of **10H₂** in 91% (111 mg) and **11H₂** in 5.0% (6 mg) yields. **10H₂**: *R_f* (silica-CH₂Cl₂) = 0.75; ¹H NMR (400 MHz, CDCl₃): δ 15.7 (s, 1H, exchangeable with D₂O), 8.79 (t, ³*J* = 4.0 Hz, 2H), 8.67–8.59 (m, 4H), 8.17 (d, ³*J* = 8.0 Hz, 4H), 7.99 (t, ³*J* = 8.0 Hz, 4H), 7.81–7.66 (m, 12H), –2.19 (s, 1H, exchangeable with D₂O), –2.33 (s, 1H, exchangeable with D₂O) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 188.2, 151.8, 145.7, 141.5, 141.4, 141.0, 139.9, 138.9, 138.0, 134.6, 134.5, 134.0, 133.6, 132.8, 129.0, 128.8, 128.4, 128.3, 128.0, 127.9, 127.6, 127.2, 127.1, 124.1, 122.2, 116.5, 114.1 ppm; UV-vis (CH₂Cl₂) λ_{max} (log ϵ) 405 (5.37), 460 (4.45), 614 (3.87), 667 (3.75) nm; IR (neat, diamond ATR): see Figure S3, SI; HR-MS (ESI⁺, cone voltage = 30 V, 100% CH₃CN) *m/z* calcd for C₄₄H₃₀N₅O₂ ([M·H]⁺) 660.2400, found 660.2381. **11H₂**: *R_f* (silica-CH₂Cl₂/1% MeOH) = 0.11; ¹H NMR (400 MHz, CDCl₃): δ 10.8–10.6 (br s, 1H), 8.71 (d, ³*J* = 4.0 Hz, 1H), 8.54 (s, 1H), 8.45 (d, ³*J* = 4.0 Hz, 1H), 8.14–8.12 (m, 2H), 8.03–7.98 (m, 2H), 7.78–7.70 (m, 6H), –2.09 (s, 1H, exchangeable with D₂O) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 154.7, 141.6, 141.5, 140.4, 137.2, 134.9, 134.8, 134.4, 133.9, 133.5, 128.5, 128.4, 128.2, 128.1, 127.6, 127.0, 123.4, 113.2 ppm; UV-vis (CH₂Cl₂) λ_{max} (log ϵ) 420 (5.11), 530 (4.05), 559 (sh), 610 (3.74), 661 (3.83) nm; IR (neat, diamond ATR): see Figure S12, SI; HR-MS (ESI⁺, cone voltage = 30 V, 100% CH₃CN) *m/z* calcd for C₄₄H₃₁N₆O₂ ([M·H]⁺) 675.2508, found 675.2515.

[meso-Tetraphenyl-2-hydroxyimino-3-oxoporphyrinato]nickel(II) (10Ni). Prepared as nearly black shining crystals in 92% yield (69 mg, 0.96×10^{-4} mol) from *meso*-tetraphenyl-2,3-dioxoporphyrinato]nickel(II) (**8Ni**) (73 mg, 1.04×10^{-4} mol) in pyridine (25 mL) and H₂NOH·HCl (344 mg, \sim 50 equiv) as described for **10H₂**: *R_f* (silica-CH₂Cl₂) = 0.70; ¹H NMR (500 MHz, CDCl₃): δ 15.3 (s, 1H, exchangeable with D₂O), 8.48 (d, ³*J* = 4.8 Hz, 1H), 8.45, 8.43 (two overlapping d, ³*J* = 4.9 Hz, 2H), 8.36, 8.34 (two overlapping d, ³*J* = 5.2 Hz, 2H), 8.32 (d, ³*J* = 4.9 Hz, 1H), 7.88 (d, ³*J* = 7.2 Hz, 4H), 7.65–7.61 (m, 13H), 7.59–7.53 (m, 3H) ppm; ¹³C NMR (125 MHz, CDCl₃): δ 184.2, 151.0, 146.0, 143.8, 143.5, 141.7, 140.4, 140.0, 139.4, 137.7, 134.0, 133.9, 133.0, 132.9, 132.7, 132.3, 131.4, 130.9, 130.5, 129.9, 129.3, 128.2, 128.1, 127.7, 127.6, 127.4, 127.2, 124.5, 122.5, 116.1, 111.5 ppm; UV-vis (CH₂Cl₂) λ_{max} (log ϵ) 410 (5.50), 473 (4.73), 505 (sh), \sim 560–800 (very broad band) nm; IR (neat, diamond ATR): see Figure S6, SI; MS (DART⁺, orifice voltage = 20 V, 100% CH₃CN) *m/z* calcd for C₄₄H₂₈N₅O₂Ni ([M·H]⁺) 716.1596, found 716.1606.

[meso-Tetraphenyl-2-hydroxyimino-3-oxoporphyrinato]palladium(II) (10Pd). Prepared in 77% yield (3.64×10^{-4} mol scale) from **8Pd** according to procedure described for **10H₂**: *R_f* (silica-CH₂Cl₂) = 0.82; ¹H NMR (400 MHz, CDCl₃): δ 15.9 (s, 1H, exchangeable with D₂O), 8.61 (d, ³*J* = 5.0 Hz, 1H), 8.57 (d, ³*J* = 5.0 Hz, 1H), 8.55 (d, ³*J* = 5.2 Hz, 1H), 8.53 (d, ³*J* = 5.0 Hz, 1H), 8.38 (d, ³*J* = 5.1 Hz, 1H), 8.35 (d, ³*J* = 5.0 Hz, 1H), 8.04 (d, ³*J* = 7.9 Hz, 4H), 7.85 (d, ³*J* = 8.1 Hz, 4H), 7.77–7.61 (m, 12H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 185.2, 151.7, 145.6, 143.2, 142.3, 140.9, 140.6, 140.2, 139.5, 133.8, 133.6, 133.5, 133.4, 132.9, 132.1, 132.0, 131.8, 130.8, 130.7, 130.2, 129.9, 129.7, 129.0, 128.6, 128.5, 128.4, 128.24, 128.20, 127.6, 127.4, 127.3, 127.25, 127.23, 125.1, 119.8, 115.4 ppm; UV-vis (CH₂Cl₂) λ_{max} (log ϵ) 400 (5.29), 468 (4.48), 498 (4.23), \sim 540–740 (br peak) nm; IR (neat, diamond ATR): see Figure S9, SI; MS (DART⁺, orifice voltage = 20 V, 100% CH₃CN) *m/z* calcd for C₄₄H₂₈N₅O₂Pd ([M·H]⁺) 764.1293, found 764.1272.

Quinoline-Annulated Oxoporphyrin 9H₂. Free base mono-oxime **10H₂** (20.1 mg, 3.05×10^{-5} mol) was dissolved in toluene (10.0 mL) in a round-bottom flask equipped with a magnetic stir bar. To the stirring solution was added *p*-TSA (12 mg, 6.31×10^{-5} mol), and the mixture was heated to reflux for 30 min. When the starting material was consumed (reaction control by UV-vis and TLC), Et₃N (3 drops) was added and the mixture was evaporated to dryness by rotary evaporation. The residue was taken up in CH₂Cl₂ and filtered

through a plug of silica gel. The filtrate was washed with water (2×10 mL), dried over anhydrous Na_2SO_4 , and evaporated to dryness by rotary evaporation. The residue was subjected to column chromatography (CH_2Cl_2 –1% MeOH) to yield **9H₂** in 76% yield (15 mg): R_f (silica– CH_2Cl_2) = 0.23; ^1H NMR (500 MHz, CD_2Cl_2): δ 9.04 (d, 3J = 4.5 Hz, 1H), 8.96 (d, 3J = 7.5 Hz, 1H), 8.45 (d, 3J = 8.0 Hz, 1H), 8.39 (t, 3J = 5.0 Hz, 2H), 8.25 (d, 3J = 4.5 Hz, 1H), 8.18 (t, 3J = 4.0 Hz, 2H), 8.08 (d, 3J = 7.0 Hz, 2H), 7.99 (d, 3J = 7.0 Hz, 2H), 7.85–7.69 (m, 13H), –0.67 (br s, 2H, exchangeable with D_2O) ppm; ^{13}C NMR (100 MHz, CD_2Cl_2): δ 195.6, 151.4, 145.4, 143.8, 141.3, 141.1, 139.4, 137.1, 136.1, 135.0, 134.6, 134.1, 133.3, 133.1, 133.0, 132.9, 130.5, 130.4, 130.3, 129.0, 128.7, 128.6, 128.3, 128.2, 128.0, 127.7, 125.6, 123.8, 112.9, 109.1 ppm; UV–vis (CH_2Cl_2) λ_{max} (log ϵ) 405 (5.17), 480 (4.51), 518 (4.39), 674 (4.16), 750 (4.30) nm; IR (neat, diamond ATR): see Figure S18, SI; HR-MS (ESI⁺, cone voltage = 30 V, 100% CH_3CN) m/z calcd for $\text{C}_{44}\text{H}_{28}\text{N}_5\text{O}$ ($[\text{M}\cdot\text{H}]^+$) 642.2294, found 642.2316.

[Quinoline-annulated oxoporphyrinato]nickel(II) (9Ni). Free base **9H₂** (14.8 mg, 2.30×10^{-5} mol) was dissolved in pyridine (5 mL) and added to a hot solution of $\text{Ni}(\text{CH}_3\text{CO}_2)_2 \cdot 4\text{H}_2\text{O}$ (34.4 mg, 1.38×10^{-4} mol, 6.0 equiv) in pyridine (10 mL) in a round-bottom flask equipped with a magnetic stir bar. The mixture was heated to reflux for 12 h. When the starting material was consumed (reaction control by UV–vis and TLC), the resulting mixture was allowed to cool, the solvent was reduced to dryness by rotary evaporation, and the residue was taken up in minimal CH_2Cl_2 and subjected to flash column chromatography (silica–100% CH_2Cl_2) to yield the main product **9Ni** in 98% yield (16 mg): R_f (silica– CH_2Cl_2) = 0.14; ^1H NMR (400 MHz, CDCl_3): δ 8.72 (d, 3J = 4.7 Hz, 1H), 8.41 (t, 3J = 7.5 Hz, 2H), 8.30 (d, 3J = 4.7 Hz, 1H), 8.10 (d, 3J = 5.0 Hz, 1H), 7.94 (d, 3J = 4.8 Hz, 1H), 7.89 (two overlapping d, 4.7 Hz, 2H), 7.77 (m, 5H), 7.69–7.56 (m, 12H) ppm; ^{13}C NMR (100 MHz, CDCl_3): δ 191.3, 150.6, 149.7, 145.1, 144.0, 143.3, 142.9, 140.6, 139.0, 138.9, 138.6, 137.0, 135.2, 135.1, 134.1, 133.1, 132.8, 132.7, 132.6, 131.6, 131.1, 130.6, 130.1, 129.9, 129.7, 129.2, 128.7, 128.6, 128.1, 128.0, 127.9, 127.8, 127.6, 127.2, 126.6, 116.1, 108.9 ppm; UV–vis (CH_2Cl_2) λ_{max} (log ϵ) 407 (4.89), 474 (4.39), 499 (4.31), 713 (4.05), 777 (3.99) nm; IR (neat, diamond ATR): see Figure S21, SI; MS (DART⁺, orifice voltage = 20 V, 100% CH_3CN) m/z calcd for $\text{C}_{44}\text{H}_{26}\text{N}_5\text{ONi}$ 698.1491 ($[\text{M}\cdot\text{H}]^+$), found 698.1498.

[Quinoline-annulated oxoporphyrinato]palladium(II) (9Pd). Free base **9H₂** (15.2 mg, 2.37×10^{-5} mol) was dissolved in benzonitrile (5 mL) and added to a hot solution of benzonitrile (15 mL) and PdCl_2 (8.5 mg, 4.79×10^{-5} mol, 2.0 equiv) in a round-bottom flask equipped with a magnetic stir bar and N_2 gas inlet, and the mixture was heated to reflux for 2 h. When the starting material was consumed (reaction control by UV–vis and TLC), the resulting mixture was allowed to cool, and the solvent was removed by rotary evaporation, taken up in the minimal quantity of CH_2Cl_2 , and purified by flash column chromatography (silica gel– CH_2Cl_2 /1% MeOH). The main green band was isolated to provide **9Pd** in 70% yield (13 mg): R_f (silica– CH_2Cl_2) = 0.25; ^1H NMR (400 MHz, CDCl_3): δ 9.00 (d, 3J = 4.9 Hz, 1H), 8.78 (d, 3J = 8.0 Hz, 1H), 8.49 (two overlapping d, 3J = 7.0 Hz, 4J = 1.1 Hz, 1H), 8.43 (d, 3J = 5.0 Hz, 1H), 8.18 (d, 3J = 5.1 Hz, 1H), 8.15 (s, 2H), 7.99–7.97 (m, 3H), 7.92–7.90 (two overlapping d, 3J = 6.3 Hz, 4J = 1.8 Hz, 2H), 7.80–7.63 (m, 13H) ppm; ^{13}C NMR (100 MHz, CDCl_3): δ 191.1, 150.4, 148.4, 145.2, 142.4, 141.3, 140.3, 140.1, 139.8, 138.2, 134.0, 133.9, 133.5, 133.4, 133.3, 133.1, 133.0, 132.2, 131.9, 131.5, 130.0, 129.7, 129.6, 129.4, 129.2, 128.7, 128.6, 128.1, 127.9, 127.7, 127.4, 127.2, 126.9, 117.5, 111.0 ppm; UV–vis (CH_2Cl_2) λ_{max} (log ϵ) 406 (4.79), 471 (4.07), 738 (3.11) nm; IR (neat, diamond ATR): see Figure S24, SI; MS (DART⁺, orifice voltage = 20 V, 100% CH_3CN) m/z calcd for $\text{C}_{44}\text{H}_{26}\text{N}_5\text{OPd}$ 746.1193 ($[\text{M}\cdot\text{H}]^+$), found 746.1204.

Monoquinoline-Annulated Porphyrin Quinoline N-Oxide 12H₂. Monooxime **10H₂** (12 mg, 1.74×10^{-5} mol) was dissolved in CH_2Cl_2 (10 mL) in a round-bottom flask equipped with a magnetic stir bar. DDQ (8 mg, 3.5×10^{-5} mol, 2 equiv) was added, and the mixture was stirred for 30 min. When the starting material was consumed (reaction control by TLC), the reaction mixture was filtered

through a short plug of silica gel. The filtrate was washed with water (2×10 mL), dried over anhydrous Na_2SO_4 , and evaporated to dryness by rotary evaporation. The crude product was purified on a preparative TLC plate (silica– CH_2Cl_2 /2% MeOH) to provide **12H₂** in 94% yield (11 mg): R_f (silica– CH_2Cl_2 /5% MeOH) = 0.72; ^1H NMR (500 MHz, CDCl_3): δ 9.21 (d, 3J = 5.0 Hz, 1H), 9.06 (d, 3J = 8.0 Hz, 1H), 9.00 (d, 3J = 7.5 Hz, 1H), 8.58 (d, 3J = 5.0 Hz, 1H), 8.50 (d, 3J = 5.0 Hz, 1H), 8.35–8.31 (m, 3H), 8.11 (d, 3J = 8.0 Hz, 2H), 8.04 (d, 3J = 8.0 Hz, 2H), 7.96 (t, 3J = 7.5 Hz, 1H), 7.81 (d, 3J = 8.0 Hz, 2H), 7.78–7.70 (m, 10H), –0.05 (br s, 2H, exchangeable with D_2O) ppm; ^{13}C NMR (125 MHz, CDCl_3): δ 184.6, 155.6, 153.7, 146.4, 144.9, 143.7, 141.2, 141.0, 140.8, 138.7, 138.1, 136.3, 135.7, 134.3, 134.1, 133.7, 133.4, 132.4, 132.3, 131.1, 130.8, 129.5, 128.4, 128.2, 127.7, 127.5, 127.3, 127.0, 126.9, 125.3, 123.3, 121.9, 114.2, 101.5 ppm; UV–vis (CH_2Cl_2) λ_{max} (log ϵ) 416 (5.35), 427 (sh), 496 (4.67), 534 (4.67), 668 (4.21), 737 (4.49) nm; IR (neat, diamond ATR): see Figure S29, SI; MS (ESI⁺, cone voltage = 30 V, 100% CH_3CN) m/z calcd for $\text{C}_{44}\text{H}_{28}\text{N}_5\text{O}_2$ ($[\text{M}\cdot\text{H}]^+$), 658.2243 found 658.2213.

[Monoquinoline-annulated porphyrinato quinoline N-oxide]nickel(II) 12Ni. Prepared and purified using a preparative TLC plate (CH_2Cl_2 /2% MeOH) in 72% yield from **10Ni** (2.20×10^{-5} mol scale) according to the general procedure for **12H₂**: R_f (silica– CH_2Cl_2 /5% MeOH) = 0.84; ^1H NMR (500 MHz, CDCl_3): δ 8.89 (d, 3J = 7.6 Hz, 3J = 1.1 Hz, 1H), 8.80 (d, 3J = 4.9 Hz, 1H), 8.46 (d, 3J = 8.0 Hz, 1H), 8.42 (d, 3J = 5.0 Hz, 1H), 8.19 (d, 3J = 4.9 Hz, 1H), 8.09 (d, 3J = 4.9 Hz, 1H), 8.04 (d, 3J = 4.9 Hz, 1H), 8.00 (d, 3J = 4.8 Hz, 1H), 7.87 (t, 3J = 7.2 Hz, 1H), 7.81–7.78 (m, 4H), 7.70 (t, 3J = 7.0 Hz, 1H), 7.66–7.56 (m, 11H) ppm; ^{13}C NMR (125 MHz, CDCl_3): δ 181.5, 148.4, 143.4, 142.9, 142.1, 140.9, 140.3, 138.9, 138.7, 136.9, 135.4, 134.1, 133.8, 133.7, 132.7, 132.6, 132.1, 131.3, 131.2, 130.7, 130.5, 130.0, 129.3, 128.6, 128.4, 128.3, 127.8, 127.7, 127.5, 127.4, 127.3, 127.1, 126.9, 121.7, 116.8, 100.7 ppm; UV–vis (CH_2Cl_2) λ_{max} (log ϵ) 420 (5.14), 493 (4.59), ~580–860 (br peak) nm; MS (DART⁺, orifice voltage = 20 V, 100% CH_3CN) m/z calcd for $\text{C}_{44}\text{H}_{26}\text{N}_5\text{O}_2\text{Ni}$ 714.1440 ($[\text{M}\cdot\text{H}]^+$), found 714.1457.

[Monoquinoline-annulated porphyrinato quinoline N-oxide]palladium(II) 12Pd. Prepared and purified using a preparative TLC plate (CH_2Cl_2 /2% MeOH) in 84% yield from **10Pd** (2.67×10^{-5} mol scale) according to the general procedure for **12H₂**: R_f (silica– CH_2Cl_2 /5% MeOH) = 0.70; ^1H NMR (400 MHz, CDCl_3): δ 9.00 (d, 3J = 5.0 Hz, 1H), 8.88 (dd, 3J = 7.4 Hz, 4J = 1.0 Hz, 1H), 8.71 (d, 3J = 8.2 Hz, 1H), 8.53 (d, 3J = 5.0 Hz, 1H), 8.28 (d, 3J = 4.8 Hz, 2H), 8.23 (d, 3J = 4.9 Hz, 1H), 8.10 (d, 3J = 4.9 Hz, 1H), 8.10 (two overlapping d, 3J = 6.3 Hz, 4J = 1.7 Hz, 2H), 7.95 (two overlapping d, 3J = 6.3 Hz, 4J = 1.6 Hz, 2H), 7.81 (t, 3J = 7.0 Hz, 1H), 7.75–7.63 (m, 12H) ppm; UV–vis (CH_2Cl_2) λ_{max} (log ϵ) 416 (5.12), 489 (4.44), 664 (sh), 718 (4.19) nm; IR (neat, diamond ATR): see Figure S33, SI; MS (DART⁺, orifice voltage = 20 V, 100% CH_3CN) m/z calcd for $\text{C}_{44}\text{H}_{26}\text{N}_5\text{O}_2\text{Pd}$ 762.1121 ($[\text{M}\cdot\text{H}]^+$), found 762.1137.

Monoquinoline-Annulated Oxoporphyrin Oxime 14. Either monoquinoline-annulated oxoporphyrin **9H₂** (51.8 mg, 8.1×10^{-5} mmol) (path B, Scheme 3) or the corresponding N-oxide **12H₂** (60 mg, 9.1×10^{-5} mmol) (path C, Scheme 3) was dissolved in pyridine (20 mL) in a round-bottom flask equipped with a magnetic stir bar and N_2 gas inlet. Solid $\text{H}_2\text{NOH}\cdot\text{HCl}$ (80 mg) was added, and the mixture was heated to reflux for 2 h. When the starting material was consumed (reaction control by TLC), the reaction mixture was allowed to cool, the solvent was removed by rotary evaporation and dissolved in some CH_2Cl_2 (~10 mL), and the mixture was filtered through a glass frit (M). The volume of the filtrate was reduced and loaded onto a flash chromatography column (silica– CH_2Cl_2 /petroleum ether 3:1). Product **14H₂** was isolated in 86% (46 mg, path B) or 90% (54 mg, path C) yield. Alternatively, the reaction can be performed at ambient temperature for 36 h with comparable yields R_f (silica– CH_2Cl_2) = 0.35; ^1H NMR (400 MHz, CDCl_3): δ 15.9 (s, 1H, exchangeable with D_2O), 9.42 (d, 3J = 8.0 Hz, 1H), 9.27 (d, 3J = 4.0 Hz, 1H), 8.50–8.46 (m, 3H), 8.38 (d, 3J = 4.0 Hz, 1H), 8.32 (d, 3J = 4.0 Hz, 1H), 8.26 (d, 3J = 4.0 Hz, 1H), 8.14–8.12 (m, 2H), 8.05–8.02 (m, 2H), 7.97 (t, 3J = 8.0 Hz, 1H), 7.94–7.91 (m, 2H), 7.86 (t, 3J = 8.0 Hz, 1H), 7.77–7.67 (m, 10H), 0.23 (s, 1H, exchangeable with

D₂O), 0.18 (s, 1H, exchangeable with D₂O) ppm; the limited solubility of this compound prevented the recording of high-quality ¹³C NMR spectra; UV-vis (CH₂Cl₂) λ_{max} (log ε) 406 (sh), 417 (5.19), 461 (sh), 489 (sh), 574 (sh), 635 (sh), 659 (4.37), 700 (4.43) nm; IR (neat, diamond ATR): see Figure S39, SI; MS (ESI⁺, cone voltage = 30 V, 100% CH₃CN) *m/z* calcd for C₄₄H₂₉N₆O ([M·H]⁺) 657.2403, found 657.2414.

meso-Tetraphenyloxadiazaeporphyrin 13H₂ and Bisquinoline-Annulated Porphyrin 7H₂. Path A (Scheme 3): Bisoxime 11H₂ (13.3 mg, 1.97 × 10⁻⁵ mol) was dissolved in toluene (10 mL) in a round-bottom flask equipped with a magnetic stir bar, *p*-TSA (7.5 mg, 4.0 × 10⁻⁵ mol, ~2.0 equiv) was added, and the mixture was heated to reflux for 30 min. When the starting material was consumed (reaction control by TLC), Et₃N (2 drops) was added and the mixture was evaporated to dryness by rotary evaporation. The residue was taken up in CH₂Cl₂ and filtered through a plug of silica gel. The filtrate was washed with water (2 × 10 mL), dried over anhydrous Na₂SO₄, and evaporated to dryness by rotary evaporation. The residue was separated on a preparative TLC plate (silica-CH₂Cl₂/petroleum ether 1:1) to furnish brown 9H₂ in 66% (8.3 mg), purple 13H₂ in 19% (2.5 mg), and green 7H₂ in 12% (1.5 mg) yields. Path E (Scheme 3): Bisquinoline-annulated porphyrin 7H₂ was prepared in 64% isolated yield from monofused oxime 14 using the procedure described in path A. Only minimal hydrolysis of the oxime and no *meso*-tetraphenyloxadiazaeporphyrin 13H₂ was observed. 13H₂: *R_f* (silica-CH₂Cl₂) = 0.86; ¹H NMR (400 MHz, CDCl₃): δ 8.95 (d, ³*J* = 4.0 Hz, 1H), 8.86 (d, ³*J* = 4.0 Hz, 1H), 8.71 (s, 1H), 8.21 (d, ³*J* = 8.0 Hz, 2H), 8.16 (d, ³*J* = 8.0 Hz, 2H), 7.89–7.74 (m, 6H), -2.72 (s, 1H, exchangeable with D₂O) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 163.8, 155.4, 141.7, 140.8, 139.1, 138.0, 137.5, 134.7, 134.6, 133.4, 129.1, 129.0, 128.3, 128.0, 127.8, 127.1, 123.2, 117.3 ppm; UV-vis (CH₂Cl₂) λ_{max} (log ε) 418 (5.48), 524 (4.27), 586 (4.13), 588 (4.08), 641 (3.60) nm; IR (neat, diamond ATR): see Figure S37, SI; MS (ESI⁺, cone voltage = 30 V, 100% CH₃CN) *m/z* calcd for C₄₄H₂₉N₆O ([M·H]⁺) 657.2403, found 657.2413. 7H₂ (the spectroscopic data corresponds well to the data reported by Jeandon and Ruppert and is included here for comparison):¹⁷ *R_f* (silica-CH₂Cl₂/1% MeOH) 0.30; ¹H NMR (400 MHz, CD₂Cl₂, 25 °C): δ 8.89 (br s, 1H), 8.71–8.69 (m, 1H), 8.28 (br s, 1H), 7.90 (d, two overlapping doublets, ³*J* = 4.0 Hz, 2H), 7.83 (d, ³*J* = 8.0 Hz, 2H), 7.71–7.67 (m, 4H), 7.40 (d, ³*J* = 4.0 Hz, 2H), 7.30 (br s, 1H), 1.43 (br s, 1H) ppm; see Jeandon and Ruppert for high-temperature data; limited solubility at rt does not allow the recording of high-quality ¹³C NMR spectra; UV-vis (CH₂Cl₂) λ_{max} (log ε) 392 (4.60), 432 (sh), 513 (3.77), 571 (sh), 619 (3.95), 675 (4.03), 775 (3.79) nm; IR (neat, diamond ATR): see Figure S41, SI; MS (ESI⁺, cone voltage = 30 V, 100% CH₃CN) *m/z* calcd for C₄₄H₂₇N₆ ([M·H]⁺) 639.2292, found 639.2294.

[Bisquinoline-annulated porphyrinato]nickel(II) (7Ni). Free base 7H₂ (11.5 mg, 1.8 × 10⁻⁵ mol) or free base 15H₂ (11.4 mg, 1.7 × 10⁻⁵ mol) (path D, Scheme 3) was dissolved in pyridine (5 mL) and added to a hot solution of pyridine (10 mL) and Ni(CH₃COO)₂·4H₂O (27 mg, 1.08 × 10⁻⁴ mol, 6.0 equiv) in a round-bottom flask equipped with a magnetic stir bar. The mixture was heated to reflux for 30 min. When the starting material was consumed (reaction control by UV-vis and TLC), the resulting mixture was allowed to cool, and the solvent was removed by rotary evaporation and taken up in minimal CH₂Cl₂. The crude product was separated on a preparative TLC plate (CH₂Cl₂/5% MeOH) to furnish green 7Ni in 91% (11.7 mg, path F) or 55% (7.1 mg, path D) yields, respectively. The spectroscopic data corresponds well to the data reported by Jeandon and Ruppert and is included here for comparison:¹⁷ *R_f* (silica-CH₂Cl₂/2% MeOH) = 0.19; ¹H NMR (500 MHz, CDCl₃): δ 8.65 (d, 7.5 Hz, 1H), 8.62 (d, 7.5 Hz, 1H), 8.58 (d, ³*J* = 4.5 Hz, 1H), 8.03 (d, ³*J* = 4.5 Hz, 1H), 7.82–7.77 (m, 2H), 7.50 (d, 2H), 7.64 (s, 1H) 7.62–7.58 (m, 3H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 151.1, 147.5, 145.0, 143.9, 139.1, 138.8, 135.5, 132.7, 132.6, 131.9, 130.9, 129.9, 129.6, 128.7, 127.9, 127.7, 127.6, 126.5, 124.8, 109.6 ppm; UV-vis (CH₂Cl₂) λ_{max} (log ε) 402 (4.52), 473 (4.42), 691 (3.91), 727 (4.00), 765 (4.05) nm; IR (neat, diamond ATR): see Figure S44, SI; MS (ESI⁺, cone voltage =

30 V, 100% CH₃CN) *m/z* calcd for C₄₄H₂₅N₆Ni ([M·H]⁺) 695.1489, found 695.1488.

[Bisquinoline-annulated porphyrinato]zinc(II) (7Zn). Bisquinoline porphyrin 7H₂ (11.5 mg, 1.8 × 10⁻⁵ mol) was dissolved in warm chloroform (10 mL) in a round-bottom flask equipped with a magnetic stir bar. A warm solution of Zn(OAc)₂ (3 equiv) in MeOH (1 mL) was added and the mixture heated to reflux for 1 h. When the starting material was consumed (reaction control by UV-vis and TLC), the mixture was diluted with MeOH (15 mL) and allowed to cool to ambient temperature, and the precipitated product was filtered off and dried to yield 7Zn in 79% (10 mg) yield: *R_f* (silica-CH₂Cl₂/2% MeOH) = 0.54; ¹H NMR (400 MHz, DMSO-*d*₆): δ 9.15 (d, ³*J* = 7.6 Hz, 1H), 8.74 (m, ³*J* = 11.0 Hz, 2H), 8.01–7.89 (m, 5H), 7.73 (s, 3H), 7.54 (s, 1H) ppm; high quality ¹³C NMR spectra could not be obtained because of the limited solubility of this complex; UV-vis (DMSO) λ_{max} (log ε) 329 (4.21), 409 (4.40), 473 (4.21), 687 (3.80), 756 (4.14) nm; MS (ESI⁺, 100% CH₃CN, TOF) *m/z* calcd for C₄₄H₂₅N₆Zn ([M·H]⁺) 701.1432, found 701.1448.

[Bisquinoline-annulated porphyrinato]palladium(II) (7Pd). Free base bisquinoline porphyrin 7H₂ (11.8 mg, 1.8 × 10⁻⁵ mol) was dissolved in pyridine (2.7 mL) in a thick-walled 2 cm OD glass tube equipped with a magnetic stir bar. Pd(acac)₂ (16.4 mg, 5.4 × 10⁻⁵ mol, 3 equiv) was added, and the vessel was sealed and placed in a microwave cavity. Using an initial microwave power of 300 W, the contents of the reaction vessel were heated to 180 °C where it was held for 15 min. Upon completion, the vessel was cooled to ambient temperature, the solvent was evaporated to dryness using rotary evaporation, and the residue was separated on a preparative TLC plate (CH₂Cl₂/2% MeOH) to provide 7Pd in 55% yield (7.5 mg): *R_f* (silica-CH₂Cl₂/2% MeOH) = 0.38; ¹H NMR (400 MHz, CD₂Cl₂): δ 8.28 (d, ³*J* = 7.7 Hz, 1H), 8.19 (d, ³*J* = 7.6, ⁴*J* = 1.1 Hz, 1H), 8.09 (d, ³*J* = 4.8 Hz, 1H), 7.73–7.58 (m, 9H) ppm; ¹³C NMR (100 MHz, CD₂Cl₂): δ 149.9, 144.7, 143.8, 140.7, 139.5, 137.0, 133.6, 133.0, 131.9, 131.4, 129.8, 129.7, 128.3, 128.2, 127.0, 126.9, 126.0, 125.5, 124.5, 109.2 ppm; UV-vis (CH₂Cl₂) λ_{max} (log ε) 323 (4.43), 401 (4.64), 472 (4.48), 668 (4.00), 710 (4.22) nm; IR (neat, diamond ATR): see Figure S48, SI; MS (ESI⁺, 100% CH₃CN, TOF) *m/z* calcd for C₄₄H₂₅N₆Pd ([M·H]⁺), 743.1191 found 743.1184.

Bisquinoline-Annulated Porphyrin Quinoline *N*-Oxide 15H₂. Prepared in 96% isolated yields (20.8 mg) from monofused oxime 14H₂ (22.3 mg, 3.31 × 10⁻⁴ mmol) dissolved in CH₂Cl₂ (10 mL) and DDQ (16.9 mg, 7.44 × 10⁻⁵ mol, ~2.3 equiv) as described for *N*-oxide 12: *R_f* (silica-CH₂Cl₂/10% MeOH) = 0.78; ¹H NMR (400 MHz, CDCl₃): δ 9.20 (d, ³*J* = 8.0 Hz, 1H), 8.98–8.93 (m, 2H), 8.89 (d, ³*J* = 8.0 Hz, 1H), 8.77 (br s, 2H), 8.15 (br s, 1H), 8.10 (d, ³*J* = 4.0 Hz, 1H), 7.95–7.82 (m, 10H), 7.75–7.65 (m, 6H), 1.79 (br s, 2H, exchangeable with D₂O) ppm; ¹³C NMR (100 MHz, CDCl₃): δ 145.1, 140.3, 139.6, 133.6, 133.5, 132.2, 131.2, 130.7, 129.6, 128.8, 128.7, 128.5, 127.9, 127.5, 127.4, 126.9, 126.5, 126.0, 125.5, 125.2, 121.2, 102.6 ppm; UV-vis (CH₂Cl₂) λ_{max} (log ε) 412 (4.80), 439 (sh), 485 (sh), 521 (4.12), 568 (3.66), 638 (3.77), 703 (4.14), 779 (4.44) nm; IR (neat, diamond ATR): see Figure S51, SI; MS (ESI⁺, cone voltage = 30 V, 100% CH₃CN) *m/z* calcd for C₄₄H₂₇N₆O ([M·H]⁺) 655.2241, found 655.2221.

■ ASSOCIATED CONTENT

Supporting Information

¹H and ¹³C NMR and IR spectra of all compounds obtained and experimental details for the crystal structure determinations of 10Ni, 12Ni, 7Pd, and 15H₂, including X-ray data (CIF). This material is available free of charge via the Internet at <http://pubs.acs.org>.

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Notes

The authors declare no competing financial interest.

■ ACKNOWLEDGMENTS

This work was supported by the US National Science Foundation (CHE-0517782 and CHE-1058846). The ApexII X-ray diffractometer was funded by NSF Grant 0087210, Ohio Board of Regents Grant CAP-491, and Youngstown State University. The Prospector X-ray diffractometer was funded by NSF Grant 1337296. We thank Mathias Senge, Trinity College Dublin, for technical advice on the skeletal deviation plots.

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